Pulse Crop Health Initiative Funded Projects – Fiscal Year 2019

Breeding Projects

MP3: More protein, more peas, more profit

Clare Coyne (PI), USDA-ARS, Pullman, WA Rebecca McGee, USDA-ARS, Pullman, WA John Miller, Montana State University, Conrad, MT

Project Start Date: FY2018

Goal: To increase seed protein concentrations in yellow pea cultivars.

<u>Objectives:</u> (1) Determine the genetic variation of protein and mineral nutrient concentration in current cultivars and advanced breeding lines of yellow pea (Genotype x Environment analysis); (2) Identify single nucleotide polymorphisms associated with alleles controlling seed protein concentration in pea using genome wide association studies (GWAS); and (3) Develop and validate breeder friendly markers for increasing seed protein and mineral nutrient concentrations in new yellow pea cultivars.

Development of efficient, genotype-independent gene-editing systems for common bean and chickpea

Shawn Kaeppler (PI), University of Wisconsin, Madison, WI

Project Start Date: FY2018

<u>Goal</u>: To develop efficient methods for transformation and gene-editing in common bean and chickpea and to utilize these methods to edit health-related traits of interest.

<u>Objectives:</u> (1) Develop a transformation system to deliver gene-editing machinery; and (2) Develop an efficient gene-editing process.

Enhancing the nutritional and functional traits of dry bean through metabolomics, genetics, and breeding

Phillip McClean (PI), North Dakota State University, Fargo, ND Juan Osorno, North Dakota State University, Fargo, ND Karen Cichy, USDA-ARS, East Lansing, MI James Harnly, USDA-ARS, Beltsville, MD Phillip Miklas, USDA-ARS, Prosser, WA

Project Start Date: FY2018

<u>Goal</u>: To correlate specific dry bean metabolite patterns with important functional and agronomic traits, to identify genetic factors associated with metabolites that distinguish functional traits in different U.S. dry bean market classes, and to develop improved dry bean germplasm that combine important functional traits with sustainable agronomic performance across diverse environments.

<u>Objectives:</u> (1) Obtain metabolite fingerprints for a large number (n=300) of advanced breeding lines from the major US market classes of dry beans; (2) Identify genetic factors associated with the major metabolite compounds identified in Objective 1; (3) Perform field trials at five locations (three in WA and two in ND) with advanced breeding lines representing the major Middle American market

classes and collect performance data; and (4) Correlate specific metabolite fingerprints associated with good end use quality characteristics.

Improved short season cowpeas and development of unmanned aerial system (UAS) and other phenotyping tools to advance pulse breeding

Seth Murray (PI), Texas A&M University, College Station, TX Bir B Singh, Texas A&M University, College Station, TX

Project Start Date: FY2019

<u>Goal</u>: To breed and release new varieties of short season cowpea (black-eyed peas) for producers to profitably incorporate pulses into existing production systems and to develop new high-throughput trait analysis tools to make future breeding, selection, and management of pulse crops more efficient.

<u>Objectives:</u> (1) Determine the yield potential and composition of new short season elite cowpea breeding lines in multiple Texas environments (Genotype x Environment analysis); (2) Advance a breeding pipeline of new second cycle cowpea lines; (3) Evaluate the ability of unmanned aerial systems (UAS) as high-throughput phenotyping tools to estimate biomass, growth, and plant populations of cowpea and other pulses, towards predicting yields in breeding pipelines; and (4) Develop near infrared spectroscopy (NIR) to estimate and predict composition, including protein, fiber, and minerals, of cowpeas and other pulses.

Improving the nutritional value of chickpeas

George Vandemark (PI), USDA-ARS, Pullman, WA Dilrukshi Thavarajah, Clemson University, Clemson, SC

Project Start Date: FY2018

<u>Goal</u>: To develop improved chickpea varieties with superior agronomic traits and nutritional qualities.

<u>Objectives:</u> (1) Assess field-grown chickpea breeding lines and varieties for seed concentrations of iron, zinc, fiber, protein, and fatty acids; (2) Identify genetic markers associated with seed concentrations of protein, iron, zinc, and fatty acids; and (3) Use diversity panel accessions with desirable nutritional qualities as parents in crosses with commercial varieties and adapted USDA-ARS breeding lines to develop improved germplasm and varieties.

Sustainability Projects

Increasing nitrogen fixation potential in pulses for environmental and economic sustainability

Clain Jones (PI), Montana State University, Bozeman, MT Kevin McPhee, Perry Miller, Scott Powell, Montana State University, Bozeman, MT Maryse Bourgault, Montana State University, Havre, MT

Project Start Date: FY2018

<u>Goal</u>: To enhance nitrogen fixation of pulse crops and increase residual nitrate in the soil for subsequent crops, thereby increasing grain yield, protein, and net economic returns.

<u>Objectives:</u> (1) Assess nitrogen fixation and residual nitrate for a range of pea and lentil lines and cultivars grown in the Northern Plains to allow pulse breeders to identify and breed for higher nitrogen fixing varieties; (2) Determine the nitrogen fixation response in lentils from starter potassium plus sulfur fertilizer with and without inoculant; and (3) Evaluate the effects of granular inoculant, peat powder seed coat inoculant, and double inoculant (granular plus peat powder, peat powder at 2x rate) on pea nitrogen fixation, residual nitrate-nitrogen, yield, and protein.

Optimizing nodulation in chickpea for enhanced nitrogen fixation

Audrey Kalil (PI), North Dakota State University, Williston Research Extension Center, Williston, ND Nonoy Bandillo, North Dakota State University, Fargo, ND

Project Start Date: FY2019

<u>Goal</u>: To improve nodulation and nitrogen fixation in chickpea using diverse rhizobia from the Northern Great Plains and to identify chickpea varieties with superior nitrogen fixing capabilities.

<u>Objectives:</u> (1) Identify rhizobia strains with improved abiotic stress tolerance, nodulation, and N fixation for use as chickpea inoculants; (2) Determine best practices regarding chickpea seed treatment and fertilization for maintaining nodulation and N fixation; (3) Identify breeding lines and existing varieties that have superior nodulation and N fixing capabilities; and (4) Determine variability in host gene expression and underlying genes responsible for enhanced nodulation and nitrogen fixation in chickpea.

Field experiments to incorporate pulse crops in cropping systems and assess soil health and plant water use efficiency

Zachary Kayler (PI), University of Idaho, Moscow, ID Xi Liang, University of Idaho, Aberdeen, ID

Project Start Date: FY2019

<u>Goal</u>: To quantify the potential of lentils, chickpeas, or dry peas in rotation or intercropped with barley on soil health, pulse performance, and barley production and to determine pulse responses to water stress to improve management practices in non-irrigated agronomic systems.

<u>Objectives:</u> (1) Determine the soil health and plant physiology responses of lentils, chickpeas, and dry peas grown in rotation and intercropped with barley; (2) Determine short term and seasonal carbon allocation to seeds, roots, and stems; (3) Evaluate the effect of including pulses on barley production; and (4) Assess the impact of water stress on pulse-barley production and on soil health indicators (e.g., organic matter and microbial community).

Using native rhizobia to improve salt-tolerance in field pea

Christopher Graham (PI), South Dakota State University, Rapid City, SD Sen Subramanian, South Dakota State University, Brookings, SD

Project Start Date: FY2019

<u>Goals:</u> To develop a multi-species (field pea and rhizobia) system that is resistant to increased soil salinity in the Northern Great Plains.

<u>Objectives:</u> (1) Increase the sustainability of field pea production in the Northern Great Plains by creating a more resilient plant-microbiome system resistant to increasing soil salinity; (2) Identify candidate field pea genomes with tolerance to increasing soil salinity; (3) Identify candidate native

rhizobia with tolerance to increasing soil salinity; (4) Compare and contrast the efficacy of native rhizobia with commercial inoculants under abiotic stress; and (5) Test the efficacy of different inoculant delivery systems (granular, peat, liquid) from both native rhizobia and commercial inoculants.

Sustainable field pea cropping systems for the Great Plains

Kraig Roozeboom (PI), Kansas State University, Manhattan, KS Ignacio Ciampitti, Zach Stewart, Kansas State University, Manhattan, KS Lucas Haag, Kansas State University, Colby, KS Augustine Obour, Kansas State University, Hays, KS John Holman, Kansas State University, Garden City, KS

Project Start Date: FY2018

<u>Goal</u>: To develop sustainable pea cropping systems for the central Great Plains.

<u>Objectives:</u> (1) Determine the relative productivity of spring and winter pea grown across a range of environments and cropping systems in Kansas; (2) Determine relative differences in nitrogen fixation and net nitrogen input to the system between spring and winter pea when grown in Kansas; (3) Evaluate the effect of including peas in Kansas rotations on soil health indicators; and (4) Evaluate the effect of incorporating peas on the small-grain segment of Kansas crop rotations.

Sustainability and health impact assessment of US pulses

Greg Thoma (PI), University of Arkansas, Fayetteville, AR Naomi Fukagawa, USDA-ARS, Beltsville, MD Peter Arbuckle, USDA-ARS, Beltsville, MD

Project Start Date: FY2018

<u>Goal</u>: To enhance the sustainability of the global food supply through optimized production of pulses, and to discover and promote the health and nutritional benefits of regular pulse consumption.

<u>Objectives:</u> (1) Establish national-scale benchmark of environmental sustainability across multiple dimensions (climate change, eutrophication, etc.) for the production and consumption of major pulse crops; (2) Submit lifecycle inventory data sets to the National Agricultural Library Digital Commons; (3) Evaluate the environmental sustainability of alternative diets with varying quantities of pulses, working in conjunction with USDA nutritionists to ensure that each alternative diet is nutritionally equivalent; and (4) Evaluate the environmental sustainability of alternative pulse production management systems, including a consequential lifecycle analysis paradigm focused on the production and health effects of pulses.

Food Technology Projects

Optimization in the production of protein hydrolysates from chickpea as novel functional food ingredients in the prevention of type-2 diabetes

Elvira de Mejia (PI), University of Illinois, Urbana, IL Brian Jacobsen, University of Illinois, Urbana, IL

Project Start Date: FY2019

<u>Goal</u>: To optimize the digestion conditions of chickpea proteins to compare their antidiabetes potential and to characterize the peptide composition of chickpea protein hydrolysates to generate ingredients with health benefits suitable in foods.

<u>Objectives:</u> (1) Optimize the processing conditions to generate protein hydrolysates with antidiabetic potential from chickpea proteins as well as to generate major peptides produced from chickpea varieties with specific amino acid sequences and predicted bioactivity; (2) Evaluate the techno-functional properties of chickpea protein hydrolysates and their incorporation in a baked snack food matrix; and (3) Determine the effect and mechanism of action of a chickpea protein hydrolysate in preventing type-2 diabetes using a murine model of diet-induced metabolic dysfunction.

Tailoring processing strategies to produce the new generation of chickpea proteins and prebiotic oligosaccharides

Juliana Maria Leite de Moura Bell (PI), University of California, Davis, California Daniela Barile, David Mills, University of California, Davis, California

Project Start Date: FY2019

<u>Goal</u>: To use an advanced mass spectrometry guided extraction process to produce chickpea protein concentrates (proteins and carbohydrates) with improved functionality and to convert chickpea processing byproducts (fiber and hulls) into selective prebiotic oligosaccharides.

<u>Objectives:</u> (1) Produce chickpea protein concentrates with increased protein content and functionality, along with diverse oligosaccharide composition, through the use of the aqueous (AEP) and enzyme-assisted aqueous extraction process (EAEP); (2) Convert chickpea hulls and the insoluble fiber generated by the AEP/EAEP of chickpea flour (byproduct) into prebiotic oligosaccharides using a modified Fenton reaction; and (3) Evaluate prebiotic and anti-infective actions of the skim fractions (proteins/peptides + carbohydrates) produced during the AEP/EAEP processing, and of the bioactive oligosaccharides produced by the conversion of chickpea hulls and insoluble fraction.

Impact of storage on functionality and nutritional and phytochemical compositions of pea, lentil, and chickpea

Clifford Hall (PI), South Dakota State University, Brookings, SD Atanu Biswas, USDA-ARS National Center for Agricultural Utilization Research, Peoria, IL

Project Start Date: FY2019

<u>Goals</u>: To understand how storage practices affect the functionality and nutritional composition of pulses and to provide the food industry with guidance in the handling of pulses during long-term storage.

<u>Objectives:</u> (1) Assess the flour, protein and starch functionality of pulses stored under (a) ambient conditions typical of North Dakota, Montana and the Palouse region for up to 4 years and (b) combinations of several relative humidities and temperatures typical of environments where pulses are exported; (2) Determine the nutrient and phytochemical compositions of the stored pulses; (3) Assess the functionality and nutrient/phytochemical composition of different pulse cultivars stored under harsh storage conditions identified in Objective 1; and (4) Characterize the impact of pulse storage on the application of flour in crackers and cookies.

Flavor, nutrition and functional properties of pea protein

Baraem (Pam) Ismail (PI), University of Minnesota, St. Paul, MN

Project Start Date: FY2018

<u>Goal</u>: To characterize the flavor, functionality, nutritional quality, and physiological properties of pea protein subjected to targeted isolation and enzymatic modification and to identify ways to increase pea protein's success in replacing partially or wholly traditional proteins in various food applications.

<u>Objectives:</u> (1) Produce pea protein isolates (PPI) and hydrolysates (PPH) and determine their structure, molecular interactions, surface properties, and functionality; (2) Identify the aroma and taste compounds in PPI and PPH that provide an undesirable flavor, and elucidate formation pathways to guide protein isolation and processing protocols; and (3) Determine the protein digestibility corrected amino acid score (PDCAAS) of PPI and PPH and evaluate their potential to promote a reduction in adiposity using an animal model.

Starch inclusion complexes to enable new uses for starch-rich byproducts of pulse processing

James Kenar (PI), USDA-ARS, Peoria, IL Frederick Felker, USDA-ARS, Peoria, IL

Project Start Date: FY2019

<u>Goal</u>: To enable the pulse food industry to increase the utilization of starch-rich pulse fractions from dry peas, lentils, chickpeas, and dry beans in food products.

<u>Objectives:</u> (1) Enable methods to prepare complex-containing pulse materials containing starch host-guest complexes by jet cooking starch-rich pulse fractions in combination with flavor, nutraceutical, or other functional guest molecules; (2) Characterize and determine bulk physicochemical and functional properties of the modified pulse materials; and (3) Develop food product prototypes using the modified pulse materials and evaluate functional and sensory properties.

The effect of food processing on fermentable oligosaccharides from pulse crops in human colon and its microbiota

Sean Liu (PI), USDA-ARS, Peoria, IL Mukti Singh, USDA-ARS, Peoria, IL Devin Rose, University of Nebraska, Lincoln, NE

Project Start Date: FY2018

<u>Goal</u>: To examine the effect of various methods of processing of pulses on digestive health and barriers to pulse consumption.

<u>Objectives:</u> (1) Assess food processing methods with pulse crops that yield lower amounts of fermentable oligosaccharides and that maintain or enhance beneficial microbiota in the colon; and (2) Conduct *in vitro* and *in vivo* studies of fermentable oligosaccharides and changes in colonic microbiota.

Optimizing pulse protein functionality

Brennan Smith (PI), University of Idaho, Moscow, ID Girish Ganjyal, Washington State University, Pullman, WA

Project Start Date: FY2018

<u>Goal</u>: To increase the functionality of pulse protein isolates through optimization of processing and biochemical modifications while maintaining or improving nutritional quality.

<u>Objectives:</u> (1) Determine the biochemical composition of starting pulse materials; (2) Optimize extraction protocols for maximum protein solubility; (3) Determine the effects of glucose addition and deamidation on the functional properties of pulse protein isolates; (4) Determine the effects of ultra-sonication and chemical disulfide bond cleavage on pulse protein isolate functionality; and (5) Determine the nutritional quality of the experimental treatments through *in vitro* digestibility studies.

Human Health Projects

Hidden nutrition: Understanding the encapsulation dynamics of the cotyledon cell to optimize consumer acceptability and nutritional benefits of dry beans

Karen Cichy (PI), USDA-ARS, East Lansing, MI Ray Glahn, USDA-ARS, Ithaca, NY Donna Winham, Iowa State University, Ames, IA

Project Start Date: FY2018

<u>Goal</u>: To determine how bean seed cell wall differences achieved via genetic diversity or processing methodology influence nutrient bioavailability, consumer acceptability, and glycemic response of dry beans.

<u>Objectives:</u> (1) Characterize the composition of the cotyledon cell wall in a unique set of dry bean germplasm with diverse cooking times and culinary attributes; (2) Evaluate the flour milling quality in dry bean germplasm with diverse cooking times and measure the nutrient bioavailability and consumer acceptability in whole beans and foods made with bean flours; and (3) Test the glycemic response of whole beans vs. foods made with bean flours.

Pulse resistant starch: Interplay between processing, the microbiome and health

Darrel Cockburn (PI), The Pennsylvania State University, University Park, PA

Project Start Date: FY2019

<u>Goal</u>: To examine the factors that influence health benefits received from pulse resistant starches as mediated by the gut microbiome

<u>Objectives:</u> (1) Investigate the influence of resistant starch from different pulses, processing steps, and dietary history using *in vitro* fecal fermentation studies to measure microbial diversity and butyrate production; and (2) Investigate the influence of resistant starch from different pulses, processing steps, and dietary history using defined microbial fermentations to measure microbial diversity and versity and butyrate production.

Understanding the pulse-gut relationship and its role in modifying systemic inflammation and insulin sensitivity in humans

Indika Edirisinghe (PI), Illinois Institute of Technology, Bedford Park, IL Amandeep Sandhu, Britt Burton-Freeman, Illinois Institute of Technology, Bedford Park, IL

Project Start Date: FY2019

<u>Goal</u>: To determine the role of pulse food consumption in a healthy diet on key health endpoints associated with the prevention of diabetes mellitus and cardiovascular disease development in at risk populations.

<u>Objectives:</u> (1) Characterize indices of systemic inflammation and gut microbiota composition and function after chronic (12 weeks) intake of pulses compared to control diet in human over weight/obese – insulin resistant (OW/OW-IR) participants; and (2) Characterize dietary- and microbial-derived metabolite pools after regular intake of pulses (12 weeks) in human OW/OB-IR participants compared to control diet.

Gut microbiota dependent and independent impacts of dietary pulses on pre- and postprandial metabolism and inflammation in overweight/obese humans

Mary Miles (PI), Montana State University, Bozeman, MT

Brian Bothner, Carl Yeoman, Seth Walk, Collen McMilin, Wan-Yuan Kuo, Tan Tran, Montana State University, Bozeman, MT

Project Start Date: FY2019

<u>Goal:</u> To determine gut microbiome dependent and independent impacts of pulse consumption on metabolic resilience and metabolic risk profiles for type 2 diabetes and cardiovascular disease risk.

<u>Objectives:</u> (1) Determine the impact of pulse (green lentil and black bean) consumption on postprandial triglyceride and inflammation responses to a high-fat meal challenge; (2) Determine the extent to which the gut microbiome and changes in the gut microbiome induced by pulse consumption influence health impacts; and (3) Measure metabolomic profiles to elucidate underlying mechanisms linking pulse consumption to improved health.

Comparative analysis of chickpea, dry pea, lentil, and dry bean for human health traits

Henry Thompson (PI), Colorado State University, Fort Collins, CO Adam Heuberger, Colorado State University, Fort Collins, CO Pamela Wolfe, Wolfe Statistical Consulting LLC, Santa Fe, NM Tiffany Weir, Colorado State University, Fort Collins, CO

Project Start Date: FY2018

<u>Goal</u>: To compare the anti-obesogenic activity of low and high dietary fiber cultivars of dry bean, chickpea, dry pea, and lentil and to assess how these pulses affect histological and molecular characteristics associated with gut health and functional activity of the gut associated microbiome

<u>Objectives:</u> (1) Determine how energy balance and lipid metabolism are impacted by low- and highdietary fiber cultivars of chickpea, dry bean, dry pea, and lentil; (2) Determine how pulse consumption affects histologic and molecular characteristics associated with gut health; and (3) Determine whether differences exist in nutrient and small molecule profiles among pulse crops and across low- versus high-dietary fiber cultivars within a given pulse crop.

Mechanisms of dry bean mediated anti-obesogenic activity

Henry Thompson (PI), Colorado State University, Fort Collins, CO Corey Broeckling, Colorado State University, Fort Collins, CO Pamela Wolfe, Wolfe Statistical Consulting LLC, Santa Fe, NM

Project Start Date: FY2018

<u>Goal</u>: To identify the components of beans responsible for health benefits, to screen and improve bean germplasm for value-added health traits, and to develop biomarkers for use in the clinical evaluation of health benefits arising from bean consumption.

<u>Objectives:</u> (1) Determine how fat deposition is partitioned in bean-fed versus control-fed mice that are provided isocaloric amounts of diet; (2) Evaluate the extent to which bean consumption affects caloric uptake and the fraction of ingested energy that is excreted in the feces, using a mouse model; (3) Use indirect calorimetry to determine how bean consumption affects respiratory quotient and/or energy expenditure; (4) Examine the role of bean consumption on the activation of AMP-activated protein kinase and its effect on lipid metabolism; and (5) Examine functional changes in the gut microbiome mediated by bean consumption, focusing on bile salt hydrolase activity and how this affects farnesoid X receptor (FXR) activity in the ileum.